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Interspecific interactions alter functionality and promote the keystone species in a synthetic four-species community

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Introduction

- **Biofilms** are highly diverse, harbouring multiple, interacting species. Such interspecies interactions lead to emergent properties unique to the community setting impacting composition, stability and functionality [1].
- **Biofilm formation** is an important process for many soil bacterial species in natural environments to colonize plant roots, interact with plants, and ultimately affect plant growth and health [2][3].

Results

> Four-species community enhanced plant drought tolerance when inoculated coculture compared to monocultures [4]



- Here, we aimed to study the how interspecific interactions shaped community dynamics and functionality of a synthetic four-species biofilm community.
- We studied a four-species consortium (SPMX), composed of Stenotrophomonas rhizophila (Sr), Xanthomonas retroflexus (Xr), Microbacterium oxydans (Mo) and Paenibacillus amylolyticus (Pa), previously shown to interact synergistically in biofilm formation. We used Arabidopsis thaliana plants as hosts to evaluate bacterial impacts. The bacterial community was studied using molecular analyses (quantitative PCR and amplicon sequencing) and fluorescence in situ hybridization (FISH-CLSM) for analysis of spatial organization.
- The study's **hypotheses** are that (1) the four strains form a biofilm on the root, retain the water and protect the plant from drought, and (2) emergent properties within the four stains impact bacterial root colonization and functionality.



> Comparison of root colonization by the four-species co-culture and monocultures [5]



Fig.2. (A) Root colonization by SPMX co-culture and monocultures, captured by confocal laser scanning microscopy (CLSM) at five days (D5); the bacterial colonization was visualized by staining with SYTO9; the root was stained with calcofluor white (CFW) (scale bar = 50 µm); (B) and (C) quantification of bacterial colonization on the roots by 3D quantitative analysis and colony forming units (CFU); data were normalized by average root length (mm); (D) SPMX biofilm formation visualized by FISH on the root surfaces.

Spatial organization and host impacts [5]



> P. amylolyticus affected spatial organization of the four-species biofilm community on plant roots [5]



Fig.3. (A) Differences in spatial organization of the four-(SPMX) and three-species (SMX) biofilm formed on the roots over time to 15 days. (B) biomass quantification (µm³, bio-volume) of each species colonizing the roots throughout the SPMX and SMX biofilm at D5, D10, and D15.

Functional differences between four- and three-species biofilm communities on plant growth over time [5]







Stenotrophomonas rhizophila (Sr) Xanthomonas retroflexus (Xr)

Days post co-cultivation (DPC)



p = 0.45

p = 0.66

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Fig.4. (A) Differences in growth phenotype of plants inoculated with four-species (SPMX) and three-species (SMX) co-cultures with removal of P. amylolyticus over time up to 15 days. (Scale bar = 1 cm) (B) main root length (cm) and shoot fresh weight (mg) of Arabidopsis seedlings cocultivated with either four-species (SPMX) or three-species (SMX) co-cultures at D5, D10, and D15 (n = 9); data are based on nine biological replicates (n = 9) from three independent experiments.

References:

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[3] Liu, Yunpeng, et al. "Root colonization by beneficial rhizobacteria." *FEMS Microbiology Reviews* 48.1 (2024). [4] Yang, Nan, et al. "Emergent bacterial community properties induce enhanced drought tolerance in Arabidopsis." NPJ biofilms and microbiomes (2021).

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Conclusion

Our results highlight the significance of emergent properties in multispecies biofilm communities for enhancing plant drought tolerance. Moreover, interspecific interactions are essential for the establishment of the keystone species, shaping community function to promote plant growth. Such bacterial interactions should be considered in the design of synthetic communities for biotechnological application.







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